



EVALUATION OF *NIGELLA SATIVA* SEEDS ON BROILER CHICKS HEMATOLOGICAL, BLOOD BIOCHEMICAL PARAMETERS AND ANTIOXIDANT ENZYMES**S. S. A. Hassan**

Anim. and Poult. Prod. Dep., Fac. of Agric., Damanhour Uni., Egypt

Corresponding Author: Saber S. Hassan; E-mail: saber.shihata77@gmail.com

Received: 23/05/2021

Accepted: 08/08/2021

ABSTRACT: The present study targets to look deep at the influence of *Nigella sativa* seeds (NSS) supplementation on hematological, blood biochemical measurements, liver and kidney functions and antioxidant enzymes of Arbor Acres broiler chicks. A total of 140 unsexed one day old Arbor Acres broiler chicks were randomly divided into four treatment groups during the period from 1-35 days of age. The chicks were randomly assigned in a straight run experimental design among four treatments, each replicated 7 times with 5 unsexed chicks per replicate. The 1st group was fed a commercial basal diet without any supplementation (control), the 2nd, 3rd and 4th groups were fed control diet supplemented with NSS at 0.5, 1.0 and 1.5% levels. Chicks fed basal diet and basal diet without supplementation or with 1.5% NSS had significantly higher RBCs than the 0.5 and 1% NSS supplemented groups. Chicks fed the NSS at 0.5% showed a higher plasma albumin level than the control group, 1 and 1.5 NSS groups. Feeding diet supplemented with 1% NSS had significantly ($P \leq 0.05$) achieved the lowest triglyceride and cholesterol compared with chicks fed diet that contained 0.5 and 1.5% NS seeds groups. Significantly ($P \leq 0.05$) higher high-density lipoprotein (HDL) was recorded in chick fed diet that contained 1.5% NSS compared with the control and the other treatments. Chicks fed diet that contained 1% NSS had significantly ($P \leq 0.05$) achieved a lower plasma aspartate amino transferase (AST). All of supplementation with different NSS levels had a significantly ($P \leq 0.05$) higher plasma creatinine and lower plasma urea to creatinine ratio than the control group. Chicks fed diet contained 0.5% NSS had significantly ($P \leq 0.05$) achieved a higher plasma superoxide dismutase and glutathione peroxidase activity compared with the control, 1 and 1.5% NSS supplemented groups. Chicks fed the diet containing 1% NSS showed higher plasma glutathione level than the control group, 0.5 and 1.5% NSS groups. Feeding diet with supplemented 0.5 and 1.5% NSS were significantly ($P \leq 0.05$) increased plasma glutathione reductase and catalase compared with the control and 1% NSS groups.

Conclusion: These findings indicated that NSS had improved lipid profile, liver function and antioxidant profile of broiler chicks. Thus, 1% NSS is considered safe due to having no acute toxic side effects as reported through the experimental period.

Key words: Broiler; *Nigella sativa* seeds; liver and kidney functions; antioxidant.

INTRODUCTION

Poultry industry has advanced in several focus points for instance; feeding, genetics, immunity enhancement and improved management for a higher productivity, better growth performance and more meat yield (Gunal *et al.*, 2006). Modern poultry farming is under a continuous demand to provide for high quality meat and egg simultaneously with a lower price under the condition of doing without antibiotics and other therapeutic utilization in animals' diets.

Medicinal plants and their derivatives are known to be safe and efficient substitute for animal production industry because of their positive influence on different physiological responses in poultry as they provide a wide variety of activities for the biological systems in terms of immunity, endocrine gland, digestive tract and a wide range of other body systems El-kashef (2020). The black seed is one of those medicinal plants which is also known by the name "black cumin" (*Nigella sativa* L.). In the Islamic literature (hadith) the Prophet Mohammed (Peace Be Upon Him (PBUH)) had described the healing capacity of the black seed by saying "Keep on the using of this black seed because it possesses a cure for every disease except for death" (Bukhari, 2018). *Nigella sativa* is a popular herbal plant that has been commonly used to work out various health issues for about 2000 years (Darakhshan *et al.*, 2015). NSS also consists of significant levels of minerals such as calcium, iron, zinc, copper, phosphorus, niacin, thiamine, pyridoxine, and folic acid (Takruri and Dameh, 1998; Ramadan, 2007).

Moreover, the phytochemical analyses of NSS has demonstrated that they contain over a hundred phytoconstituents, predominantly alkaloids, saponins, sterols and essential oils, nonetheless, the structure of all of them has not been chemically realized nor biologically confirmed (Mamun and Absar, 2018; Ghahramanloo *et al.*, 2017). Furthermore, they possessed a number of biologically active constituents, for instance; dithymoquinone, thymol, thymoquinone, which are known to be a pharmaceutically active substances of NSS, the most primarily bioactive component that the black seed possesses is the essential oil in the range of 27.8-57% (Ali and Blunden 2003), alpha-hedrin, carvacrol, nigellidine and nigellicine-N-oxide (Al-Homidan *et al.*, 2002; Nasir *et al.*, 2005). The composition of NSS makes it yield to many properties that are demonstrated in figure (1) (Badary *et al.*, 2003; Khader *et al.*, 2010; Mohideen *et al.*, 2003; Srinivasan, 2018).

In clinical trials, *Nigella sativa* showed to have an effect when included as an alternative for the traditional hypolipidemic and antidiabetic medicines. Inhibiting dietary cholesterol absorption suppressed hepatic cholesterol production and up-regulation of LDL receptors leading to the lipid-reducing effects of *Nigella sativa*. Some studies and researches pointed out that NSS plays a role in the antibacterial activity (El-Kamali *et al.*, 1998; Mouhajir *et al.*, 1999; Nair *et al.*, 2005), where the NSS and their abundant extracts have an effect on gram-positive and gram-negative bacteria and suppress the production of

Broiler; *Nigella sativa* seeds; liver and kidney functions; antioxidant.

aflatoxin (Nasir and Grashorn, 2006), and have many biological characteristics such as antiparasitic (Mahmoud *et al.*, 2002). The effects of dietary *Nigella sativa* meal (NSM) or oils on poultry performance have been investigated and it has been concluded decisively that the consumption of feed, body weight and efficient performance in broilers are unquestionably influenced. (Halle *et al.*, 1999; Tollba and Hassan, 2003; Guler *et al.*, 2006; Ziad *et al.*, 2008; AL-Beitawi *et al.*, 2009; Erenner *et al.*, 2010; Toghyani *et al.*, 2010). Some studies showed that diets which contained 10% of NSM demonstrated no effects on growth performance (Al-Homidan *et al.*, 2002). Consequently, the current study targeted the investigation of the effect of the supplementation of NSS on hematological, blood biochemical parameters, liver and renal functions and antioxidant enzymes of broiler chicks.

MATERIALS AND METHODS

The study was conducted at the Poultry Research Unit at El-Bostan Farm, animal and poultry department, Faculty of Agriculture, Damanhour University approved the experiment. This study was carried out to investigate the effectiveness of dietary inclusion of NSS on blood hematological, biochemical parameters, liver and renal functions and antioxidant enzymes of broiler chickens.

***Nigella sativa* seeds sources**

The NSS was purchased commercially from the local commercial market at Damanhour city, Egypt.

Chicks, diets and experimental design

One hundred and forty (140) unsexed one day old Arbor Acres broiler chicks were fed the experimental diets (Table 1) during the period from 1-35 days of age. The chicks were randomly assigned in a straight run experimental design among

four treatments, each replicated 7 times with 5 unsexed chicks per replicate. Corn-soybean meal basal diet was formulated to be isocaloric and isonitrogenous to meet the nutritional requirements according to NRC (1994). The basal diet was fed without supplements (control group) or supplemented with NSS levels of 0.5, 1.0 and 1.5% (Table 1). NSS contain crude protein, crude fiber, ether extract, moisture and ash (21.16%, 10.92%, 31.97%, 5.58% and 1.88%), respectively according Mohammed (2016).

Husbandry of chickens:

Chicks were kept in battery brooders (40×45×60 cm) under similar managerial and hygienic conditions in a semi-open house. Water and mash diets were given *ad libitum* throughout the whole experimental period. The brooding temperatures were 33, 31 and 30°C during the 1st, 2nd and 3rd weeks of age, respectively. During 21-35 days of age, the average ambient temperature and relative humidity (RH %) were 30 ± 3°C and 45 ± 4%, respectively. The lighting regimen was 23:1 light-dark cycle.

Data Collection:

Blood samples collection

At 35 days of age, six chicks from each group were randomly taken at 08:00-09:00 am and about 3 ml blood was collected from the wing vein into vacuoliner tubes with K3-EDTA (1 mg/mL).

Non-coagulated blood was divided into two parts. The first part was used, shortly after collection, for estimating blood picture, whereas, the second part was centrifuged at 4000 rpm for 15 minutes and the clear plasma was separated and stored in a deep freezer at -20°C until biochemical analysis. All blood

biochemical variables were determined calorimetrically using commercial kits.

Hematological parameters

Red blood cells' count (RBCs $10^6/\text{ml}^3$) was done according to Feldman *et al.* (2000). Hemoglobin (Hb) concentration (g/dl) and the percentage of packed cells volume (PCV %) were measured according to Drew *et al.* (2004). The average volume (size) of RBC (MCV, μm^3) = [hematocrit (%) / RBC] \times 10, the average weight of hemoglobin in RBC (MCH, pg) = [hemoglobin concentration (g/dL) / RBC] \times 10 and the average concentration of hemoglobin in the RBC (MCHC, %) = [hemoglobin (g/dL) / hematocrit (%)] \times 100 counts were calculated. A thin blood film was prepared by using a small drop of blood. The blood film was completely dried before staining using Giemsa stain.

Blood biochemical parameters

Protein profile

To evaluate the changes in the protein profile at 35 days of age in chickens, plasma total protein (g/dl) and albumin (g/dl) were determined using special kits delivered from Diamond diagnostics (23 EL-Montazah St. Heliopolis, Cairo, Egypt, <http://www.diamonddiagnostics.com>) by means of spectrophotometer (Beckman DU-530, Germany) according to guidelines of Armstrong and Carr (1964) and Doumas *et al.*, (1977), respectively. Plasma globulin level (g/dl) was calculated by the difference between total protein and albumin, since the fibrinogen usually comprises a negligible fraction (Sturkie, 1986). Albumin to globulin ratio was also calculated. Albumin to globulin ratio (A/G) was calculated by dividing the total

level of albumin of the total level of globulin.

Lipid profile

Plasma total lipid, triglyceride concentrations (mg/dl) were determined in blood plasma using special kits delivered from Diamond diagnostics (23 EL-Montazah St. Heliopolis, Cairo, Egypt, <http://www.diamonddiagnostics.com>) by means of spectrophotometer according to recommendation of (Fringes *et al.*, 1972). Plasma total cholesterol (mg/dl) was determined on individual bases using the specific kits according to recommendation of (Bogin and Keller, 1987). Plasma samples were analyzed for high-density lipoprotein (HDL, mg/dl), low-density lipoprotein (LDL, mg/dl) and HDL to LDL ratio (HDL/LDL) was calculated by dividing the total level of HDL of the total level of LDL.

Liver function

The transaminase enzymes activities of plasma aspartate amino transferase (AST) and plasma alanine amino transferase (ALT), as U/L were determined by calorimetric method of (Reitman and Frankel, 1957) and AST to ALT ratio was calculated. Alkaline phosphatase (ALP, U/L) concentration was determined according to the colorimetric method of (Bauer, 1982).

Kidney function

Renal function, urea (mg/dl) and creatinine (g/dl) were assessed in the serum based on the suggestions of Bartles *et al.*, (1972) and Sampson *et al.*, (1980), respectively, and the urea-to-creatinine ratio was calculated.

Oxidative status

superoxide dismutase (SOD, IU), glutathione (GSH, mg/dl), glutathione

Broiler; *Nigella sativa* seeds; liver and kidney functions; antioxidant.

peroxidase activity (GPx, U/ml) and glutathione reductase (GRx, U/ml) and were determined according to Koracevic *et al.* (2001), Chiu *et al.* (1976), Beutler *et al.* (1963) and Misra and Fridovich (1972), respectively. Catalase (nmol/min/ml) was determined according to Rindler *et al.*, (2013). Total antioxidant capacity (TAC, nmol/l) was determined according to Koracevic *et al.* (2001). Malondialdehyde (MDA, $\mu\text{mol/L}$) was determined according to Richard *et al.* (1992).

Statistical analyses:

Analysis of variance was done using a one-way analysis of variance, as described by SAS[®] (2009), using the following model:

$$Y_{ij} = \mu + F_i + e_{ij}$$

Where: Y=the dependent variable, μ = the overall mean; F_i = the effect of NSS treatments and e_{ij} = the random error. The replicate was the experimental unit. The mean difference at $P \leq 0.05$ was tested using Tukey's HSD test.

RESULTS AND DISCUSSION

Effect of *Nigella sativa* seeds on blood hematological

The effect of supplementation of NSS on hematological parameters of broiler chicks is presented in table (2). Chicks fed basal diet and basal diet supplemented with 1.5% NSS had significantly higher RBC's than the 0.5 and 1% NSS supplemented groups. No significant differences were observed between the control group and all of levels of NSS treatment groups for Hgb, PCV, MCV, MCH and MCHC.

In accordance with the current results, the positive effect of *Nigella sativa* seeds on hematology could be linked to the highly active components, particularly thymoquinone and thymohydroquinone, as they have a robust antioxidation activity

(Arslan *et al.*, 2005) and thus, the increased RBC counts in NSS treated quail chicks may be attributed to the decreased lipid peroxide in RBC membrane bringing on a decreased sensitivity of RBC to hemolysis (Toghyani *et al.*, 2010). Shokrollahi and Sharifi (2018) noticed that RBC counts showed a significant improve in the 1% and 1.5% NSS treatments ($P \leq 0.05$). There were no big variations in Hb and PCV levels among the treatments ($P > 0.05$). In addition to that, Toghyani *et al.* (2010) discovered that NSS were the cause of the increased RBC, Hb and PCV levels in broilers. Mixed outcomes on the effect of NSS on the hematological profile of healthy broiler are available (Al-Mansour *et al.*, 2011; Ghasemi *et al.*, 2014). Haqa *et al.*, (2020) who quoted that including NSS in the diet showed no major effect on RBC count and Hb concentration in broiler chicks.

Effect of *Nigella sativa* seeds on plasma proteins and lipid profile

The results in table (3) showed the effect of supplementation of NS on plasma proteins and lipid profile of broiler chicks. The results indicated that chicks fed diet that contained 1.5% NSS had significantly ($P \leq 0.05$) achieved a lower plasma total protein compared with the control and 0.5% NSS supplemented groups. The highest plasma total protein observed in chicks fed basal diet. Chicks fed the NSS at 0.5% showed higher plasma albumin level than the control, 1 and 1.5% NSS groups. Feeding diet supplemented with 1% NSS significantly ($P \leq 0.05$) increased plasma total lipid compared with the control and the other treatment (0.5 and 1.5% NSS) groups. Chicks fed diet containing 1% NSS had significantly ($P \leq 0.05$) achieved the lowest triglyceride and cholesterol

compared with chicks fed diet containing 0.5 and 1.5% NSS groups. The highest triglyceride was observed in birds fed diet containing 0.5% NSS. Significantly ($P \leq 0.05$) higher HDL was recorded in chick fed diet containing 1.5% NSS compared with the control and the other treatments. While, the lowest HDL observed was in the control group. Feeding diet with supplementation of 1% NSS significantly ($P \leq 0.05$) decreased LDL compared with the control and 0.5% NSS groups. Chicks fed diet containing 1.5% NSS presented a higher HDL to LDL ratio than the other groups and the lowest level of HDL to LDL ratio was observed with the control group.

Likewise, Attia and Al-Harhi (2015) revealed that the inclusion of *Nigella sativa* oil at 0.5 and 1.5g/kg in the diet significantly lessened the overall plasma total protein and albumin. While, Hassan *et al.*, (2007) noticed an improvement of serum albumin with the supplementation of NSS. Blood serum proteins are a sign for the state of the body and the changes that occur under the effect of internal and external factors (Toghyani *et al.*, 2010). However, Miraghaee *et al.* (2011) concluded that NSS had no obvious impact on serum protein, yet increased the levels of serum albumin in broiler chicks. The average Plasma protein, albumin and globulin of Hubbard broiler chicks fed on 4 g/kg NSS were approximately alike those of the control group (El-Ghamry *et al.*, 2002). Shokrollahi and Sharifi (2018) noticed no major contrast in the levels of albumin and total protein in the respective treated groups ($P > 0.05$). El-kashef (2020) demonstrated that the total protein was significantly improved in groups 3% and

6% NSM in comparison with those treated with 9% NSM or to the control group and higher albumin levels were observed in the groups receiving 3% and 6% NSM. AboSaleh *et al.*, (2019) and Kumar *et al.* (2017b) discovered that the concentration of total protein tended to be higher in the NSS than in the control group, which had a positive correlation with higher NSS doses (Kumar *et al.*, 2017a). Elevated serum total protein with NSS supplementation was noted by Al-Beitawi and El-Ghousein (2008). Yattoo *et al.* (2012) discovered a significantly higher serum total protein and albumin with 1% NSS supplementation as compared to the control group, and Khan *et al.* (2012) noticed enhanced total protein value compared with the 1.25% antibiotic or non-supplemented diet groups. Tollba and Hassan (2003) and Hassan *et al.* (2007) discovered increased serum total protein, albumin and globulin in birds fed diets with high levels of NSS. Saleh (2014) noted that feeding NS oil at the level of (1 ml/kg) elevated serum total protein and albumin concentrations, but not globulin concentration, and albumin to globulin ratio compared to the control group (Kumar *et al.*, 2017a). The improvement in albumin level may be ascribed to that NSS possess an immune-stimulating effect. El-Ghamry *et al.* (2002) discovered an opposite finding, in which supplementing with 4 g/kg NSS in the diet showed a little effect on plasma total protein, albumin or globulin.

As stated by Aziza *et al.*, (2019), serum total protein and globulin levels were considerably higher in the group supplemented with *Nigella sativa* oil compared to the positive control group. Meanwhile, the albumin content and

Broiler; *Nigella sativa* seeds; liver and kidney functions; antioxidant.

albumin/globulin ratio did not indicate any substantial difference among all experimental categories (in quail). Soliman *et al.*, (2017) demonstrated a highly significant improvement ($p < 0.01$) in serum total protein and albumin and a highly significant decline ($p < 0.01$) in globulin level in broiler chicks fed 5.6% *N. sativa* Linn.

Current results demonstrated that cholesterol and triglycerides` levels were reduced in layer chicks supplemented with 1% NSS. In accordance with our results Akhtar *et al.* (2003) noted that the serum total cholesterol and triglycerides were less in birds fed on a diet containing 1% NSS in than a diet NSS-free (Hermes *et al.*, 2011; El-Bagir *et al.*, 2006; Hassan *et al.*, (2007); Al-Beitawi and El-Ghousein (2008); AL-Beitawi *et al.*, 2009; Ghasemi *et al.*, 2014; Yattoo *et al.* (2012). Shokrollahi and Sharifi (2018) noticed that the levels of cholesterol and triglycerides declined significantly ($P < 0.05$) in the groups treated with 1 and 1.5% NSS compared to control one, respectively. LDL concentration declined significantly ($P < 0.05$) in NSS treated broiler chicks compared to the control ones. The levels of HDL showed no significant differences among the treatments ($P > 0.05$); however, HDL concentration was higher in the treated broiler chicks. Concentrations of cholesterol are likely to be lower for NSS compared to control group, which had a negative linear correlation with increasing doses (5, 10, and 20 g/kg) of NSS (Kumar *et al.*, 2017a). To the contrary, Khalaji *et al.* (2011) demonstrated that 1% NSS in broiler diets had no effect on the serum plasma cholesterol (Khalaji *et al.*, 2011). The decrease in serum cholesterol levels might be ascribed elevated bile secretion (El-Dakhkhny *et al.*, 2000). Brunton

(1999) proposed that the decrease in serum cholesterol may be linked to the lowering influence of thymoquinone and unsaturated fatty acids on synthesizing cholesterol by hepatocytes or the fractional reabsorption from the small intestine. Furthermore, NSS constitute considerable amount of sterols, particularly β -sitosterol that could inhibit the absorption of dietary cholesterol (Khan *et al.*, 2012). Those constituents might stimulate the excretion of cholesterol into the intestine being oxidized by bile acids (Tollba and Hassan, 2003). This reduction in serum cholesterol level of broiler chicks fed NSS diets probably proposes a general decline in lipid mobilization.

Ali *et al.*, (2014) found out that 0.50% NSS in diet resulted in a reduced level of blood LDL. Although, they also noticed that raising NSS diet up to 0.75%, resulted in an increase in the levels of HDL. El-kashef (2020) demonstrated that diets containing various levels of NSM showed a reduction in ($P \leq 0.05$) serum total cholesterol. Except for the group fed 9% NSM, all treatments showed an increase in the HDL fraction, at the same time decreasing the LDL fraction.

Animal studies have showed that NSS possess hypocholesterolemic and hypolipidemic properties (Toghyani *et al.*, 2010). In accordance with our results, cholesterol and triglycerides were reduced in chicks given NSS with the levels of 10 g NSS/kg diet (Miraghaee *et al.*, 2011; Shewita and Taha, 2011), 10 and 15 g NSS/kg diet (AL-Beitawi and EL-Ghousein, 2008; Yalçın *et al.*, 2012) led to decline in the levels of cholesterol and triglycerides. Likely, Bölükbaşı *et al.* (2009) proposed that NSS was the reason for the reduction of the levels of triglycerides. The decline in serum

triglycerides could be linked to NSS volatile oils (Thymoquinone and dithymoquinone) (Swamy and Tan, 2000). The choleric activity of NSS has the capacity to lower the levels of serum cholesterol and triglycerides (El-Dhakhny *et al.*, 2000) through either reducing the synthesis of triglycerides and cholesterol by hepatocytes or lessening its fractional reabsorption from the small intestine (Brunton, 1996). Akhtar *et al.* (2003) demonstrated that NSS has the capacity to lower concentration the serum LDL in layer hens. The amount of serum HDL changed a little among the treatments, though HDL showed no significant increase in NSS-treated chicks. 1.5% NSS showed a significant enhancement in HDL concentration (Akhtar *et al.*, 2003; AL-Beitawi and EL-Ghousein, 2008). Contrarily Tufan *et al.* (2015) declared that NSS and their oil showed no effect on the concentrations of HDL and LDL in quail chicks.

Effect of *Nigella sativa* seeds on liver and renal function indices

Table (4) illustrated the effect of supplementation of NSS on liver and renal function indices of broiler chicks. The results indicated that chicks fed diet containing 1% NSS had significantly ($P \leq 0.05$) achieved a lower plasma AST compared with chicks fed diet supplemented with 0.5 and 1.5% NSS groups. The highest plasma AST protein observed in chicks fed diet containing 0.5% NSS group. No significant ($P \leq 0.05$) differences were found among the control group and treatment groups for plasma ALT, AST to ALT ratio and alkaline phosphatase. Chicks fed the NSS 1.5% showed a higher plasma urea level than the control and chicks fed diet containing

0.5 and 1% NSS groups. All of supplementation with different NSS levels had significantly ($P \leq 0.05$) higher plasma creatinine and lower plasma urea to creatinine ratio than the control group. The scarcity of significant differences in plasma ALT among treatment diets indicates normal liver function in birds fed NSS-containing diets. Although, the decline in the activity of AST noticed in birds fed a NSS diet of 1% may propose that it has properties that may enhance liver health. Consequently, Attia and Al-Harhi (2015) demonstrated that plasma ALT in the control, *Nigella sativa* oil at 0.5 and 1g/kg of diet groups was lower than that in the 1.5g *Nigella sativa* oil/kg of diet group. The levels of AST were comparable in the monitor, 0.5, and 1.5g *Nigella sativa* oil/kg diet categories. The level of alkaline phosphatase showed a significant decrease in 1.5g *Nigella sativa* oil/kg of diet group than in the other groups. The level of plasma urea showed a significant increase in the groups fed 1.5g *Nigella sativa* oil /kg of diet than the other groups. Plasma creatinine had a different pattern, and the plasma urea/creatinine ratio was more in the control and 1.5g *Nigella sativa* oil/kg diet groups than in the 0.5 and 1g *Nigella sativa* oil/kg diet groups. Except for 9% of NSM, plasma AST and ALT declined. As the liver is shown to contain enzymes such as ALT and AST, when it is enfeebled, it secretes those enzymes into the blood stream (Kaplan *et al.*, 2003). Moreover, Soliman *et al.*, (2017) discovered that the enzymes of the liver (ALT, AST) and creatinine levels exerted a highly significant enhancement ($p < 0.01$) in broiler chicks fed on a diet containing 5.6% NSS, while urea

Broiler; *Nigella sativa* seeds; liver and kidney functions; antioxidant.

demonstrated a significant elevation in all treated groups compared to the normal control group with highly significant differences ($p < 0.01$). Liver indices (AST, ALT) and renal functions indices (urea and creatinine) may be useful in decide the health and physiological status of an animal (Toghyani *et al.*, 2010; Attia *et al.*, 2015).

Effect of *Nigella sativa* seeds on antioxidant indices

Effects of experimental treatments on antioxidant indices of broiler chicks presented in table (5). The results indicated that chicks fed diet contained 0.5% NSS had significantly ($P \leq 0.05$) achieved a higher plasma superoxide dismutase and glutathione peroxidase activity compared with the control, 1 and 1.5% NSS supplemented groups. Chicks fed the diet contained 1% NSS showed higher plasma glutathione level than the control group, 0.5 and 1.5% NSS groups. Feeding diet with supplemented 0.5 and 1.5% NSS significantly ($P \leq 0.05$) increased plasma glutathione reductase compared with the control and 1% NSS groups, the lowest glutathione reductase was observed in chicks fed diet containing 1% NSS group. Chicks fed diet containing 0.5 and 1.5% NSS had significantly ($P \leq 0.05$) increased plasma catalase compared with the control and chicks fed diet supplemented with 1% NSS groups. The lowest plasma catalase level was observed in the control group. Significantly ($P \leq 0.05$) higher plasma total antioxidant capacity was observed in chicks fed diet containing 0.5 and 1% NSS compared with the control and the chicks fed diet containing 1.5% NSS group. While, the lowest plasma total antioxidant capacity was found in the control group. There were no significant ($P \leq 0.05$) differences in plasma

malondialdehyde among the control and different levels of diet supplemented NSS groups.

NSS possess a highly antioxidative activity and has the capacity to reduce hepatic lipid peroxidation as well as enhancing the activity of many enzymes such as SOD, GSH, catalase, and adenosine deaminase, which they play an important role in reducing the oxidative stress in broilers' livers (Sogut *et al.* 2008; Azeem *et al.*, 2014). Liver indices (AST, ALT) and renal functions (urea and creatinine) could be useful in deciding on the health and physiological status of an animal (Tuluze *et al.*, 2009). The marked antioxidant activity of NSS and thymoquinone could be a realizable modern antioxidant agent to be used as nutrients for the health promotion and disease prevention (Yimer *et al.*, 2019). Recent findings showed an improvement in health status of broilers fed on diets containing various additives, as it was indicated that they have no effects on liver and renal functions and increased antioxidant enzymes.

In accordance with the current outcomes, Attia and Al-Harhi (2015) found out that the groups fed on *Nigella sativa* oil in the diets exerted a significant increase in the levels of SOD, GSH, GPx, and GRx than the control group. Treating with NSS was linked to the increased levels of GSH activity, which may be ascribed to thymoquinone, which possesses potent anti-oxidative and anti-inflammatory properties (Nili-Ahmadabadi *et al.*, 2011). Aziza *et al.*, (2019) discovered that the hepatic GSH level was significantly enhanced just in the groups supplemented with NS oil when compared to both control groups. Accordingly, in a rat model, the fixed and essential oil of NSS had a clear increase

in GR and GPx versus the oxidative stress resulted from potassium bromate (Sultan *et al.*, 2015).

Linked to the key bioactive components of black cumin essential oil, NSS could work as an antioxidant which inhibits the development of free radical and raises the antioxidant enzyme activity (Tülüce *et al.*, 2009). This result is in accordance with El-Hack *et al.*, (2018). Polyphenol-containing substances could suppress nuclear factor kappa signaling; thereby elevating the level of nuclear factor erythroid 2-related factor 2 activation which preserves the cells from oxidative sabotage and promotes antioxidant activity (Tangney and Rasmussen, 2013). As a result, the now time usage of NSS in

thirty postmenopausal women demonstrated an un questionable reduction in the levels of plasma MDA with an elevated activity in erythrocyte GPx and SOD after two months of consumption (Mostafa *et al.*, 2015).

CONCLUSION

These findings indicated that NSS had improved lipid profile (decreased triglyceride and cholesterol), liver function (lower AST) and antioxidant indices of broiler chicks. As a result, NSS is marked safe as a result that they have no acute toxic side effects like what was reported throughout the experimental period.

Broiler; *Nigella sativa* seeds; liver and kidney functions; antioxidant.

Table (1): Ingredients and compositions of the experimental diets

Ingredients	Starter				Grower diet			
	0	0.5	1	1.5	0	0.5	1	1.5
Corn	54.02	53.52	53.02	52.52	61.02	60.52	60.02	59.52
Soybean meal (44%)	33.0	32.5	32.5	32.5	26.0	26.0	26.0	26.0
Corn gluten meal (60%)	4.0	4.0	4.0	4.0	4.0	4.0	4.0	4.0
Mixed oils (%)	4.5	5.0	5.0	5.0	4.5	4.5	4.5	4.5
Nigella seeds (%)	0.0	0.5	1.0	1.5	0.0	0.5	1.0	1.5
Sodium chloride	0.3	0.3	0.3	0.3	0.3	0.3	0.3	0.3
Dl- methionine	0.28	0.28	0.28	0.28	0.28	0.28	0.28	0.28
L-lysine	0.2	0.2	0.2	0.2	0.2	0.2	0.2	0.2
Dicalcium phosphate	1.6	1.6	1.6	1.6	1.6	1.6	1.6	1.6
Limestone	1.7	1.7	1.7	1.7	1.7	1.7	1.7	1.7
Vit. and min. mixture	0.3	0.3	0.3	0.3	0.3	0.3	0.3	0.3
Antioxidant	0.1	0.1	0.1	0.1	0.1	0.1	0.1	0.1
Total	100	100	100	100	100	100	100	100
ME kcal/ kg diet ²	3200	3200	3200	3200	3200	3200	3200	3200
Dry matter ³	90.15	89.8	89.31	88.87	90.06	89.61	89.17	88.73
Crude protein	23.0	23.0	23.0	23.0	20.0	20.0	20.0	20.0
Calcium	1.0	1.0	1.0	1.0	1.0	1.0	1.0	1.0
Non-phytate phosphorus ²	0.48	0.5	0.48	0.47	0.45	0.45	0.45	0.45
Methionine+cystine ²	0.76	0.7	0.75	0.74	0.68	0.68	0.68	0.68
Lysine ²	1.36	1.3	1.34	1.34	1.17	1.17	1.17	1.17
Crude fibre ³	4.25	4.2	4.18	4.1644	3.91	3.90	3.88	3.8674
Crude fat ³	6.66	7.14	7.12	7.11	6.81	6.80	6.78	6.77

¹Vit+Min mix. Provides per kilogram of the diet: Vit. A, 12000 IU, vit. E (dl- α -tocopheryl acetate) 20 mg, menadione 2.3 mg, Vit. D3, 2200 ICU, riboflavin 5.5 mg, calcium pantothenate 12 mg, nicotinic acid 50 mg, Choline 250 mg, vit. B₁₂ 10 μ g, vit. B₆ 3 mg, thiamine 3 mg, folic acid 1 mg, d-biotin 0.05 mg. Trace mineral (mg/ kg of diet): Mn 80 Zn 60, Fe 35, Cu 8, and Selenium 0.1 mg.

²Calculated values, ³Analyzed values.

Table (2): Effect of *Nigella sativa* seeds on blood hematological of broiler chicks at 35 days of age

Item	Control	<i>Nigella sativa</i> seeds			SEM	P value
		0.5%	1%	1.5%		
RBC's (10 ⁶ /mm ³)	10.54 ^a	10.42 ^b	10.39 ^b	10.55 ^a	0.059	0.0002
Hgb (g/dl)	11.3	10.9	11.5	11.5	0.044	0.438
PCV (%)	32.8	32.3	33.3	34.1	0.104	0.296
MCV (fl/cell)	31.11	31.00	32.05	32.32	1.661	0.764
MCH (Ug)	10.72	10.46	11.06	10.90	0.626	0.135
MCHC (%)	34.4	33.7	34.5	33.7	0.155	0.188

^{a,b,c} Means within the same row with different superscript letters are significantly different at p≤0.05

RBC= Red blood cells; Hgb= Hemoglobin; PCV= packed cell volume; MCV= Mean Corpuscular volume; MCH= Mean Corpuscular Hemoglobin; MCHC= Mean Corpuscular Hemoglobin concentration

Table (3): Effect of *Nigella sativa* seeds on plasma proteins and lipid profile of broiler chicks at 35 days of age

Item	Control	<i>Nigella sativa</i> seeds			SEM	P value
		0.5%	1%	1.5%		
T.pro (g/dl)	5.49 ^a	5.40 ^{ab}	5.27 ^{bc}	5.18 ^c	0.018	0.010
Alb. (g/dl)	2.51 ^b	2.90 ^a	2.53 ^b	2.56 ^b	0.016	0.0030
Glb. (g/dl)	2.98	2.50	2.73	2.62	0.029	0.1060
A/G ratio	0.866	1.196	0.943	0.986	0.017	0.0590
T. lipid (mg/dl)	106.0 ^b	106.0 ^b	111.0 ^a	104.0 ^b	0.407	0.0400
Trig. (mg/dl)	185.0 ^{bc}	191 ^a	183 ^c	187 ^b	0.258	0.0001
Cho. (mg/dl)	209.0 ^b	213 ^a	206 ^c	212 ^a	0.32	0.0020
HDL (mg/dl)	36.2 ^c	39.0 ^b	38.1 ^b	40.8 ^a	0.177	0.0001
LDL (mg/dl)	135.8 ^a	135.8 ^a	131.3 ^b	133.8 ^{ab}	0.149	0.0020
HDL/LDL ratio	0.267 ^c	0.287 ^b	0.290 ^b	0.305 ^a	0.016	0.0030

^{a,b,c} Means within the same row with different superscript letters are significantly different at p≤0.05

T.pro= Total protein; Alb= Albumin; Glb= Globulin; A/G ratio= Albumin to Globulin ratio; T.lip= Total lipids; Trig= Triglycerides; Cho= Cholesterol; HDL= High density lipo-protein; LDL= Low density lipo-protein

Broiler; *Nigella sativa* seeds; liver and kidney functions; antioxidant.

Table (4): Effect of *Nigella sativa* seeds on liver and renal function indices of broiler chicks at 35 days of age

Item	Control	<i>Nigella sativa</i> seeds			SEM	P value
		0.5%	1%	1.5%		
Liver functions						
AST (U/L)	61.9 ^{bc}	64.1 ^a	60.9 ^c	63.4 ^{ab}	0.196	0.0001
ALT (U/L)	56.4	57.7	56.2	57.1	0.249	0.6687
AST/ALT ratio	1.10	1.11	1.08	1.11	0.005	0.0628
Alkaline phosphatase (U/L)	12.4	10.3	11.9	11.0	0.153	0.1057
Renal functions						
Urea (mg/dl)	26.6 ^b	28.0 ^b	28.3 ^b	31.7 ^a	0.232	0.0003
Creatinine (mg/dl)	0.77 ^b	1.29 ^a	1.31 ^a	1.31 ^a	0.02	0.0001
U/C ratio	35.4 ^a	21.9 ^b	22.6 ^b	24.4 ^b	0.522	0.0001

^{a,b,c} Means within the same row with different superscript letters are significantly different at $p \leq 0.05$

AST= Aspartate aminotransferase; ALT= Alanine aminotransferase; AST/ALT= Aspartate aminotransferase to Alanine aminotransferase ratio; U/C ratio= Urea to Creatinine ratio

Table (5): Effect of *Nigella sativa* seeds on antioxidant indices of broiler chicks at 35 days of age

Item	Control	<i>Nigella sativa</i> seeds			SEM	P value
		0.5%	1%	1.5%		
SOD (IU)	254 ^b	268 ^a	252 ^b	254 ^b	0.483	0.0001
GSH (mg/dl)	1003 ^b	1012 ^b	1055 ^a	1006 ^b	1.567	0.0001
GPX (U/ml)	0.407 ^b	0.471 ^a	0.411 ^b	0.411 ^b	0.003	0.0001
GR (U/ml)	29.0 ^b	30.2 ^a	27.2 ^c	30.6 ^a	0.11	0.0001
Cat (nmol/min/ml)	32.8 ^c	39.8 ^a	34.7 ^b	38.7 ^a	0.151	0.0001
TAC (nmol/l)	418 ^c	453 ^a	455 ^a	444 ^b	0.754	0.0001
MAD (μ mol/l)	1.4	1.43	1.39	1.59	0.023	0.1179

^{a,b,c} Means within the same row with different superscript letters are significantly different at $p \leq 0.05$

SOD = superoxide dismutase; GSH = glutathione; GPx = glutathione peroxidase activity; GR = glutathione reductase; Cat= Catalase; TAC= total antioxidant capacity; MDA= Malondialdehyde.

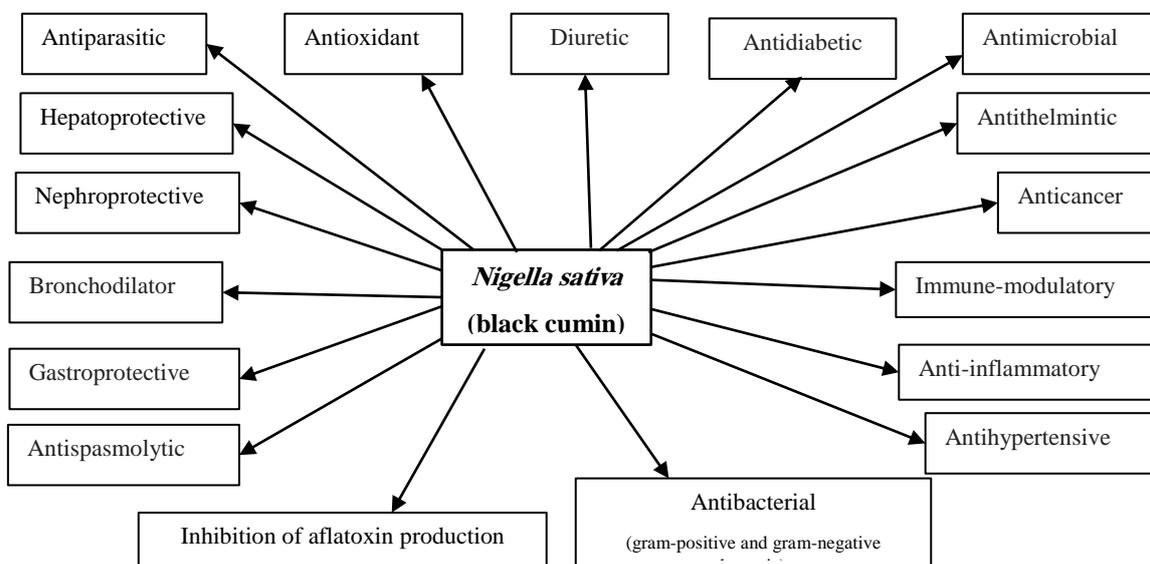


Figure (1): distinguish properties of *Nigella sativa* (black cumin)

REFERENCES

AboSaleh, Sherin; Salama, M. F.; El-Sherbini, E.; Hassan Maha, Z. 2019. The Possible Ameliorative Effect of *Nigella Sativa* on Aflatoxin-induced Liver Damage in Chicken. *A.J.V.S.* Vol. 63 (2) Oct: 113-120.

Akhtar, M. S.; Nasir, Z.; Abdur Rehman, A. 2003. Effect of feeding powdered *Nigella sativa* seeds on poultry egg production and their suitability for human consumption. *Vet. Arhiv.*, 73 (3): 181-190.

AL-Beitawi, N. A.; El-Ghousein, S. S. 2008. Effect of feeding different levels of *Nigella sativa* seeds (Black Cumin) on performance, blood constituents and carcass characteristics of broiler chicks. *International Journal of Poultry Science*, 7 (7): 715-721.

AL-Beitawi, N. A.; El-Ghousein, S. S.; Nofal, A. H. 2009. Replacing bacitracin methylene disalicylate by crushed *Nigella sativa* seeds in broiler rations and its effects on growth, blood constituents and immunity. *Livest. Sci.* 125: 304 – 307.

Al-Homidan, A.; Al-Qarawi, A. A.; Al-Waily, S. A.; Adam, S. E. I. 2002. Response of broiler chicks to dietary *Rhazya stricta* and *Nigella sativa*. *Br. Poultry Sci.* 43 (2), 291-296.

Ali, B. H.; Blunden, G. 2003. Pharmacological and toxicological properties of *Nigella sativa*. *Phytother Res*, 17, 4, 299-305.

Ali, O. A. A.; Suthama, N.; Mahfud, L. D. 2014. The effect of feeding black cumin (*Nigella Sativa*) and vitamin C on blood lipid profiles and growth performance of broilers. *Int. Refereed J. Eng. Sci.*, 3: 28-33.

Al-Mansour, S.; Al-Khalf, A.; Al-Homidan, I.; Fathi, M. M. 2011. Feed efficiency and blood hematology

Broiler; *Nigella sativa* seeds; liver and kidney functions; antioxidant.

- of broiler chicks given a diet supplemented with yeast culture. *Int. J. Poult. Sci.* 10 (8), 603–607.
- Armstrong, W. D.; Carr, C. W. 1964.** Estimation of serum total protein. In: *Physiological Chemistry Laboratory Direction*, 3rd ed. Minneapolis, Minnesota, USA: Burges Publishing Co.
- Arslan, S. O.; Gelir, E.; Armutcu, F.; Coskun, O.; Gurel, A.; Sayan, H.; Celik, I. L. 2005.** The protective effect of thymoquinone on ethanol-induced acute gastric damage in the rat *Nutrition Research*, 25, 673–680.
- Attia, Y. A.; Al-Harhi, M. A. 2015** *Nigella* seed oil as an alternative to antibiotic growth promoters for broiler chickens. *Europ. Poult. Sci.*, 79, ISSN 1612-9199.
- Attia, Y. A., Hamed, R. S.; Abd El-Hamid, A. E.; Shahba, H. A.; Bovera, F. 2015.** Effect of inulin and mannanoligosaccharides in comparison to zinc-bacitracin on growing performance, nutrient digestibility and hematological profiles of growing rabbits. *Anim. Production Science*. 55:80-86.
- Azeem, T.; Zaib-Ur-Rehman, Umar, S.; Asif, M.; Arif, M.; Rahman, A. 2014.** Effect of *Nigella sativa* on poultry health and production: a review. *Sci Lett.* 2:76–82.
- Aziza, A. E.; Abdelhamid, F. M.; Risha, E. F.; Elsayed, M. M.; Awadin, W. F. 2019.** Influence of *Nigella sativa* and rosemary oils on growth performance, biochemical, antioxidant and immunological parameters, and pathological changes in Japanese quail challenged with *Escherichia coli*. *Journal of Animal and Feed Sciences*, 28, 354–366, <https://doi.org/10.22358/jafs/114239/2019>
- Badary, O. A.; Taha, R. A.; Gamal El-Din, A. M.; AbdelWahab, M. H. 2003.** Thymoquinone Is a Potent Superoxide Anion Scavenger. *Drug Chem. Toxicol.* 26 (2): 87-98.
- Bartles, H.; Bohmer, M.; Heirli, C. 1972.** Serum creatinine determination without protein precipitation. [Article in German]. *Clin. Chem. Acta.*, 37: 193-197.
- Bauer, J. D. 1982.** Clinical laboratory methods "9th Ed.", the C.V. Company Waistline Industrial Missorri 63116 chapter 33: 555.
- Beutler, E.; Duron, O.; Kelly, B. M. 1963.** Improved method for the determination of blood glutathione. *J Lab Clin Med.* 1963; 61: 882–890.
- Bogin, E.; Keller, P. 1987.** Application of clinical biochemistry to medically relevant animal models and standardization and quality control in animal biochemistry. *J. Clin. Chem. Biochem.*, 25: 873-878.
- Bölükbaşı, S. C.; Kaynar, Ö.; Erhan, M. K.; Ürüpan, H. 2009.** Effect of feeding *Nigella sativa* oil on laying hen performance, cholesterol and some proteins ratio of egg yolk and *Escherichia coli* count in feces. *Archieve Geflügelk*, 73, 167–172.
- Brunton, L., 1996.** Agents affecting gastrointestinal water flux and motility; emesis and antiemetics; bile acids and pancreatic enzymes. *Goodman and Gillman's the Pharmacological Basis of Therapeutics*, 917-936.
- Brunton, L. L. 1999.** Agents affecting gastrointestinal water flux and motility, digestants and bile acids. *The pharmacological basis of therapeutics*.

- 8th ed. Pregman Press, Oxford, UK, pp. 914-932
- Bukhari, A. A. 2018.** Sahih-ul-Bukhari, <http://quranx.com/Search?q=Black+cumin&context=Hadith>.
- Chiu, D. T. Y.; Stults, F. H.; Tappel, A. L. 1976.** Purification and properties of rat lung soluble glutathione peroxidase. *Biochimica et Biophysica Acta*, 445: 558–566.
- Darakhshan, S.; Bidmeshki Pour, A.; Hosseinzadeh Colagar, A.; Sisakhtnezhad, S. 2015.** Thymoquinone and its therapeutic potentials. *Pharmacol Res*, 95-96, 138-158
- Doumas, B. T.; Waston, W.; Biggs, H. H. 1977.** Albumin standards and the measurements of serum albumin with bromocresol green. *Clinical Chemistry Acta*, 31: 87-96.
- Drew, P.; Charles, R. J. S.; Trevor, B.; John, L. 2004.** Oxford Handbook of Clinical Haematology. 2th Edition, Oxford University Press, USA.
- El-Bagir, N. M.; Hama, A. Y.; Hamed, R. M.; El-Rahim, A. G. A.; Beynen, A. C. 2006.** Lipid composition of egg yolk and serum in laying hens fed diets containing black cumin (*Nigella sativa*). *International Journal of Poultry Science* 5: 574-578.
- El-Dakhakhny, M.; Mahady, N. I.; Halim, M. A. 2000.** *Nigella sativa* L. oil protects against induced hepatotoxicity and improves serum lipid profile in rats. *Arzneimittel Forschung* 50: 832-836.
- El-Daly, E. S. 1998.** Protective effect of cysteine and vitamin E, *Crocus sativus* and *Nigella sativa* extracts on cisplatin-induced toxicity in rats. *J. Pharm. Belg.* 53: 87–95.
- EL-Ghamry, A. A.; EL-Mallah, G. M.; EL-Yamny, A. T. 2002.** The effect of incorporation yeast culture, *Nigella sativa* seeds and fresh garlic in broiler diets on their performance. *Egypt Poultry Science*, 22, 445–459.
- El-Hack, M. E. A.; Mahgoub, S. A.; Hussein, M. M. A.; Saadeldin, I. M. 2018.** Improving growth performance and health status of meat-type quail by supplementing the diet with black cumin cold-pressed oil as a natural alternative for antibiotics. *Environm. Sci. Pollut. Res.* 25, 1157–1167
- El-Kamali, H. H.; Ahmed, A. H.; Mohammed, A. H. 1998.** Antibacterial properties of essential oils from *Nigella sativa* seeds, *Cymbopogon citratus* leaves and *Pulicaria undulata* aerial parts. *Fitoterapia*, 69: 77 – 78.
- El-kashef, M. M. A. 2020.** Effect of different levels of *Nigella sativa* meal on growth performance, some blood biochemical and immune-responsiveness of broiler chicks. *Egypt. Poultry Sci. Vol. (40) (III):* 729-741.
- Erener, G.; Altop, N.; Ocak, H.; Aksoy, S.; Ozturk, E. 2010.** Influence of black cumin seed (*Nigella sativa* L) and seed extract on broilers performance and total coliform bacteria count. *Asian J. Anim. Vet. Adv.* 5: 128 – 135.
- Feldman, B. F.; Zinkl, J. G.; Jain, N. C. 2000.** Schalm's veterinary hematology. lippincott Williams and Wilkins, Philadelphia, USA.
- Fringes, C. S.; Fendly, T. W.; Rum, R. T.; Queen, C. A. 1972.** Improved

Broiler; *Nigella sativa* seeds; liver and kidney functions; antioxidant.

- determination of total serum lipids by the sulfo-phosphovanilin reaction. *Clinical Chemistry*; 18: 673-674.
- Ghahramanloo, K. H.; Kamalidehghan, B.; Akbari Javar, H.; Teguh Widodo, R.; Majidzadeh, K.; Noordin, M. I. 2017.** Comparative analysis of essential oil composition of Iranian and Indian *nigella sativa* L. Extracted using supercritical fluid extraction and solvent extraction. *Drug Design, Development and Therapy*, vol. 11, pp. 2221–2226.
- Ghasemi, H. A.; Kasani, N.; Taherpour, K. 2014.** Effects of black cumin seed (*Nigella sativa* L.), a probiotic, a prebiotic and a synbiotic on growth performance, immune response and blood characteristics of male broilers. *Livest. Sci.* 164, 128–134.
- Guler, T.; Dalkic, B.; Ertas, O. N.; Ciftci, M. 2006.** The effect of dietary black cumin seeds (*Nigella sativa* L.) on the performance of broilers. *Asian Aust. J. Anim. Sci.* 19 (3): 425 – 430.
- Gunal, M.; Yayli, G.; Kaya, O.; Karahan, N.; Sulak, O. 2006.** The effects of antibiotic growth promoter, probiotic or organic acid supplementation on performance, intestinal microflora and tissue of broilers. *Int. J. Poult. Sci.*, 5: 149-155.
- Halle, I.; Thomann, R.; Flachowsky, G.; Schubert, R.; Flachowsky, G.; Bitsch, R.; Jahreis, G. 1999.** Effect of ethereal (essential) oil and oilseed on the growth of broilers. *Vitamin und Zusatzstoffe in der Ernaehrung von Mensch und Tier: 7, Symposium Jena, Thuringen, Germany.*
- Haqa, I.; Hafeez, A.; Khan, R. U. 2020.** Protective effect of *Nigella sativa* and *Saccharomyces cerevisiae* on zootechnical characteristics, fecal *Escherichia coli* and hematopoietic potential in broiler infected with experimental Colibacillosis. *Livestock Science*, 239, 104119. <https://doi.org/10.1016/j.livsci.2020.104119>
- Hassan, M. S. H.; AboTaleb, A. M.; Wakwak, M. M.; Yousef, B. A. 2007.** Productive, physiological and immunological effect of using some natural feed additives in Japanese quail, Egypt. *Poult. Sci.* 27: 557- 581.
- Hermes, I. H.; Attia, F. M.; Ibrahim, K. A.; El-Nesr, S. S. 2011.** Physiological responses of broiler chickens to dietary different forms and levels of *Nigella sativa* L. during Egyptian summer season. *Journal of Agricultural and Veterinary Science* 4:17-33.
- Javed, S.; Shahid, A. A.; Haider, M. S. 2010.** Nutritional, phytochemical potential and pharmacological evaluation of *Nigella Sativa* (Kalonji) and *Trachyspermum Ammi* (Ajwain). *Journal of Medicinal Plants Research*, vol. 6, no. 5: 768-775.
- Kaplan, L. A.; Pesce, A. J.; Kazmierczak, S. C. 2003.** Liver Function. In: Sherwin, J.E. (Ed.), *Clinical Chemistry*, fourth edition. Elsevier Science, St. Louis, Toronto
- Khader, M.; Bresgen, N.; Eckl, P. M. 2010.** Antimutagenic effects of ethanolic extracts from selected Palestinian medicinal plants. *J. Ethnopharmacol.* 127(3): 319–324.
- Khalaji, S., Zaghari, M.; Hatami, S. K.; Hedari-Dastjerdi, Lotfi, L.; Nazarian, H. 2011.** Black cumin seeds, *Artemisia* leaves (*Artemisia sieberi*), and *Camellia* L. plant extract as phytogenic products in broiler diets and their effects on performance, blood constituents, immunity, and

- cecal microbial population. *Poultry Science* 90: 2500-2510.
- Koracevic, D.; Koracevic, G.; Djordjevic, V.; Andrejevic, S.; Cosic, V. 2001.** Method for the measurement of antioxidant activity in human fluids. *J. Clin. Pathol.*, 54: 356-361.
- Kumar, P.; Patra, A. K.; Mandal, G. P.; Debnath, B. C. 2017b.** Carcass characteristics, chemical and fatty acid composition, and oxidative stability of meat from broiler chickens supplemented with black cumin (*Nigella sativa*) seeds. *Animal Production Science. J. of Ani. Phy. and Ani. Nutr.* 7 February: 769 -779.
- Kumar, P.; Patra, A. K.; Mandal, G. P.; Samanta, I.; Pradhan, S. 2017a.** Effect of black cumin seeds on growth performance, nutrient utilization, immunity, gut health and nitrogen excretion in broiler chickens. *Journal of the Science of Food and Agriculture* 97: 3742-3751 doi:10.1002/jsfa.8237.
- Mahmoud, M. R.; El-Ahbar, H. S.; Saleh, S. 2002.** The effect of *Nigella sativa* oil against the liver damage induced by *Schistosoma Mansoni* infection in mice. *J. Ethnopharmacol.* 79: 1-11.
- Mamun, M. A.; Absar, N. 2018.** Major nutritional compositions of black cumin seeds cultivated in Bangladesh and the physicochemical characteristics of its oil," *International Food Research Journal*, vol. 25, no. 6, pp. 2634-2639.
- Miraghaee, S. S.; Heidary, B.; Almasi, H.; Shabani, A.; Elahi, M.; Modaber Nia, M. H. 2011.** The effects of *Nigella sativa* powder (black seed) and *Echinacea purpurea* (L.) Moench extract on performance, some blood biochemical and hematological parameters in broiler chickens. *African Journal of Biotechnology*, 19249-19254.
- Misra, H. P.; Fridovich, I. 1972.** The role of superoxide anion in the autooxidation of epinephrine and a sample assay for Superoxide dismutase. *J Biol Chem* 247: 3170-3175.
- Mohammed, H. A. 2016.** Effects of adding crushed seeds of *Nigella sativa* and leaves of *Thymus vulgaris* on chemical composition parameters of main, carcasses parts. *Int. J. Adv. Res. Biol. Sci.* 3 (12): 1-6.
- Mohideen, S.; Ilavarasan, R.; Sasikala, E.; Thirumalai, K. R. 2003.** Hepatoprotective Activity of *Nigella sativa* Linn. *Indian J. Pharm. Sci.* 65(5):550-551.
- Mostafa, R. M.; Moustafa, Y. M.; Mirghani, Z.; AlKusayer, G. M.; Moustafa, K. M. 2013.** Antioxidant effect of garlic (*Allium sativum*) and black seeds (*Nigella sativa*) in healthy postmenopausal women. *SAGE Open Medicine*, vol. 1, Article ID 2050312113517501.
- Mouhajir, F.; Pedersen, J. A.; Rejdali, M.; Towers, G. H. N. 1999.** Antimicrobial thymohydroquinones of Moroccan *Nigella Sativa* seeds detected by electron spin resonance. *Pharm. Biol.* 37: 391-395.
- Nair, M. K. M.; Vasudevan, P.; Venkitanarayanan, K. 2005.** Antibacterial effect of black seed oil on *Listeria monocytogenes*. *Food Control* 16 (5), 395-398.
- Nasir, Z.; Abid, A. R.; Hayat, Z.; Shakoor, H. I. 2005.** Effect of

Broiler; *Nigella sativa* seeds; liver and kidney functions; antioxidant.

- Kalongi (*Nigella sativa*) seeds on egg production and quality in white Leghorn layers. *J. Anim. Plant Sci.* 15: 22 - 24.
- Nasir, Z.; Grashorn, M. A. 2006.** Use of Black cumin (*Nigella sativa*) as alternative to antibiotics in poultry diets. In: M. Rodehutschord (Editor). Proceedings of 9th Tagung Schweine- und Geflügelernahrung. Halle, Saale (Germany): 210 - 213.
- Nasir, Z.; Grashorn, M. A. 2010.** Effect of Echinacea Purpurea and *Nigella Sativa* supplementation on broiler performance, carcass and meat quality. *J. Anim. Feed Sci.* 19: 94 -104.
- National Research Council, NRC (1994).** Nutrient Requirements of Poultry. 9th Ed. Nat. Acad. Press. Washington, DC., USA.
- Nili-Ahmadabadi, A., Tavakoli, F.; Hasanzadeh, G. R.; Rahimi, H. R.; Sabzevari, O. 2011.** Protective effect of pretreatment with thymoquinone against Aflatoxin B1 induced liver toxicity in mice. *Daru.* 19(4): 282.
- Ramadan, M. F. 2007.** Nutritional value, functional properties and nutraceutical applications of black cumin (*Nigella sativa* L.): an overview. *International Journal of Food Science & Technology*, vol. 42, no. 10: 1208-1218.
- Reitman, S.; Frankel, S. 1957.** A colorimetric method for the determination of serum glutamic oxalacetic and glutamic pyruvic transaminases. *Am. J. Clin. Pathol.*, 28: 56-63.
- Richard, M. J.; Portal, B.; Meo, J.; Coudray, C.; Hadjian, A.; Favier, A. 1992.** Malondialdehyde kit evaluated for determining plasma and lipoprotein fractions that react with Thiobarbituric acid. *Clinical Chemistry*, 38:704-709.
- Rindler, P. M.; Plafker, S. M.; Szweda, L. I.; Kinter, M. 2013.** High dietary fat selectively increases catalase expression within cardiac mitochondria. *J. Biol. Chem.* 288, 1979-1990.
- Sampson, E. J.; Baird, M. A.; Burtis, C. A.; Smith, E. M.; Witte, D. L.; Bayse, D. D. 1980.** A coupled-enzyme equilibrium method for urea measuring in serum. optimization and evaluation of the AACC study group on urea candidate reference method. *Clin. Chem.* 26, 816-826.
- SAS institute 2009.** SAS[®] User's Guide: Statistics. Version 5th ed., SAS Institute Inc., Cary, NC, USA.
- Shewita, R. S.; Taha, A. E. 2011.** Effect of dietary supplementation of different levels of black seed (*Nigella sativa* L.) on growth, performance, immunological, hematological and carcass parameters of broiler chicks. *World Academy of Science, Engineering and Technology* 77: 788-794.
- Shokrollahi, B.; Sharifi, B. 2018.** Effect of *Nigella sativa* seeds on growth performance, blood parameters, carcass quality and antibody production in Japanese quails. *Journal of Livestock Science (ISSN online 2277-6214)* 9: 56-64.
- Sogut, B.; Celik, I.; Tuluçe, Y. (2008).** The effects of diet supplemented with the black cumin (*Nigella sativa* L.) upon immune potential and antioxidant marker enzymes and lipid peroxidation in broiler chicks. *J. Anim. Vet. Adv.* 7:1196-1199.
- Soliman, E. S.; Hamad Rania, T.; Amira Ahmed 2017.** Prophylactic and immune modulatory influences of *Nigella sativa* Linn. in broilers exposed to biological challenge.

- Veterinary World, Vol. (10): 1447-1455: EISSN: 2231-0916.
- Srinivasan, K. 2018.** Cumin (*Cuminum cyminum*) and black cumin (*Nigella sativa*) seeds: traditional uses, chemical constituents, and nutraceutical effects. *Food Quality and Safety, Vol. 2*, 1–16 doi:10.1093/fqsafe/fyx031
- Sturkie, P. 1986.** Body fluids: Blood. P.D. Sturkie (Ed.), *Avian Physiology*, Springer-Verlag, New York, NY, pp. 103-121.
- Sultan, M. T.; Butt, M. S.; Karim, R. 2015.** *Nigella sativa* fixed and essential oil modulates glutathione redox enzymes in potassium bromate induced oxidative stress. *BMC Complementary and Alternative Medicine*, vol. 15, no. 1, article no. 330.
- Takruri, H. R. H.; Dameh, M. A. F. 1998.** Study of the nutritional value of black cumin seeds (*Nigella sativa* L.),” *Journal of the Science of Food and Agriculture*, vol. 76, no. 3: 404–410.
- Tangney, C. C.; Rasmussen, H. E. 2013.** Polyphenols, inflammation and cardiovascular disease. *Curr. Atheroscler. Rep.* 15, 324.
- Toghyani, M.; Toghyani, M.; Gheisari, A.; Ghalamkari, G.; Mohammadrezaei, M. 2010.** Growth performance, serum biochemistry and blood hematology of broiler chicks fed different levels of black seed (*Nigella sativa*) and peppermint (*Mentha piperita*). *Livest. Sci.* 129 (1-3), 173–178.
- Tollba, A. A. H.; Hassan, M. S. H. 2003.** Using some natural additives to improve physiological and productive performance of broiler chicks under high temperature conditions 2- black cumin (*Nigella Sativa*) or garlic (*Allium sativum*) Egypt. *Poult. Sci.* 23: 327 - 340.
- Tülüce, Y.; Ozkol, H.; Sogut, B.; Celik, I. 2009.** Effects of *Nigella sativa* on lipid peroxidation and reduced glutathione levels in erythrocytes of broiler chickens. *Cell Membr Free Radic Res.* 1:1–3
- Yalçın, S.; Yalçın, S.; Uzunoglu, K., Duyum, H. M.; Eltan, Ö. 2012.** Effects of dietary yeast autolysate (*Saccharomyces cerevisiae*) and black cumin seed (*Nigella sativa* L.) on performance, egg traits, some blood characteristics and antibody production of laying hens. *Livest. Sci.*, 145: 13–20.
- Yatoo, M. A.; Sharma, R. K.; Khan, A. N.; Rastogi, A.; Pathak, A. K. (2012).** Effect of fenugreek and black cumin seeds as feed additives on blood biochemical profile and performance of broilers. *Indian Journal Animal Nutrition* 29: 174-178.
- Yimer, E. M.; Tuem, K. B.; Karim, A.; Ur-Rehman, N.; Anwar, F. 2019.** *Nigella sativa* L. (Black Cumin): A Promising Natural Remedy for Wide Range of Illnesses. *Evidence-Based Complementary and Alternative Medicine*, Article ID 1528635, 16 pages. <https://doi.org/10.1155/2019/1528635>
- Ziad, H.; Abu-Dieyeh, M.; AbuDarwish, M. S. 2008.** Effect of feeding powdered black cumin seeds (*Nigella sativa* L) on growth performance of 4-8 week-old broilers. *J. Anim. Vet. Adv.* 7 (3): 86 – 290.

الملخص العربي

تقييم بذور حبة البركة على صفات كتاكيت اللحم الهيماتولوجية والبيوكيميائية في الدم والإنزيمات المضادة للأكسدة

صابر شحاته عبد الونيس حسن

قسم الإنتاج الحيواني والداخلي - كلية الزراعة - جامعة دمنهور - مصر

استهدف البحث دراسة تأثير إضافة بذور حبة البركة على صفات الدم الكيميائية والهيماتولوجية ووظائف الكبد والكلية والإنزيمات المضادة للأكسدة لكتاكيت التسمين. تم استخدام ١٤٠ كتكوتاً تسمين غير مجنسة عمر يوم حتى عمر ٣٥ يوماً. قسمت الكتاكيت عشوائياً في تصميم عشوائي بسيط إلى أربعة معاملات، بكل معاملة ٧ مكررات وبكل مكررة ٥ كتاكيت كالتالي: مجموعة الكنترول أو إضافة بذور حبة البركة بمعدل ٠.٥ و ١ و ١.٥٪ على الترتيب. زاد عدد كرات الدم الحمراء للكتاكيت التي تم تغذيتها على العليقة الكنترول أو العليقة المضافة إليها ١.٥٪ بذور حبة عن العليقة المضافة إليها بذور حبة البركة بمستويات ٠.٥ و ١٪، ارتفع مستوى ألبومين البلازما للكتاكيت التي تم تغذيتها على ٠.٥٪ بذور حبة البركة مقارنة بالكنترول و ١ و ١.٥٪ بذور حبة البركة. وكان أقل مستوى للدهون الثلاثية والكوليسترول للكتاكيت المغذاة على عليقة تحتوي على ١٪ بذور حبة البركة مقارنة بالكتاكيت التي تغذت على عليقة تحتوي على ٠.٥ و ١.٥٪ بذور حبة البركة، وأدت إضافة بذور حبة البركة بمستوى ١.٥٪ إلى زيادة الليبوبروتين عالي الكثافة مقارنة بالكنترول والمعاملات الأخرى. وانخفض مستوى الأسبارتات أمينو ترانسفيراز في بلازما الكتاكيت التي تغذت على عليقة تحتوي على ١٪ بذور حبة البركة مقارنة بالكتاكيت التي تغذت على عليقة تحتوي على ٠.٥ أو ١.٥٪ بذور حبة البركة. كل المستويات المختلفة من بذور حبة البركة أدت إلى ارتفاع كرياتينين البلازما وانخفاض نسبة اليوريا إلى الكرياتينين مقارنة بالكنترول. إضافة بذور حبة البركة بمستوى ٠.٥٪ أدت إلى زيادة أكسيد ديسموتاز البلازما ونشاط الجلوتاثيون بيروكسيديز مقارنة بالكنترول والمجموعات الأخرى. بينما زاد مستوى جلوتاثيون البلازما بإضافة بذور حبة البركة بمستوى ١٪ مقارنة بالكنترول والمستويات الأخرى. وزاد مستوى الجلوتاثيون بإضافة بذور حبة البركة بمستوى ١٪ مقارنة بالكنترول وإضافة حبة البركة بمعدل ٠.٥ و ١.٥٪. وزاد مستوى الجلوتاثيون والكاتالاز بإضافة بذور حبة البركة بمستوى ٠.٥ و ١.٥٪ مقارنة بالكنترول ومستوى ١٪ من بذور حبة البركة.

الخلاصة: أشارت النتائج إلى أن استخدام بذور حبة البركة أدت إلى تحسن تمثيل الدهون (انخفاض الدهون الثلاثية والكوليسترول)، وتحسن وظائف الكبد (انخفاض AST) وزادت مضادات الأكسدة لكتاكيت التسمين. وبالتالي تعتبر بذور حبة البركة بمعدل ١٪ آمنة لعدم وجود أي آثار جانبية سامة لها خلال فترة التجربة.

الكلمات الدالة: بدارى اللحم، بذور حبة البركة، وظائف الكبد والكلية، مضاد الأكسدة